



Production, physicochemical, microbiological and nutritional analysis of the liquor obtained from the peelings of *Ananas Comosus* reinforced by the root extract of *Mondia whitei* harvested in Benin

KOUDORO Yaya Alain^{1*}, AGBANGNAN Dossa Cokou Pascal¹, DOVONON Léonce Firmin¹, OBASSA Marus Adékoulé¹, OLAYÉ Théophile¹, AVLESSI Félicien¹, Wotto valentin Dieudonné²

¹Laboratory of Study and Research in Applied Chemistry, University of Abomey-Calavi, Polytechnic School of Abomey-Calavi, 01, P. O. Box 2009, Benin,

²Physical Chemistry Laboratory Materials Molecular Modeling

Abstract In Benin, *Ananas Comosus* co-products are often left in wild dumpsites for flies and mosquitoes and are a source of foul odour. The main objective of this study is to valorize the co-products of the *Ananas Comosus* processing industry through the formulation of the liquor reinforced with bioactive substances. To do this, the secondary metabolites of the *Mondia whitei* root were identified by staining and precipitation reactions specific to each metabolite. After the elaboration of the liquor, the organoleptic and physicochemical properties of this product were determined. The contents of phenolic compounds in the liquor were evaluated by spectrophotometric assay. The quantification of its mineral salt content was carried out by atomic absorption spectrophotometry. Finally, the microbiological quality of the formulated product was determined according to ISO standards. From the results obtained, it appears that the root of *Mondia whitei* is rich in secondary metabolites. The pH of the formulated liquor is 4.75 and its soluble solids content is 15°Brix. Total phenol concentrations; in flavonoids and in condensed tannins are respectively 11.31 mg GAE/mL; 15.07 µg QE/mL and 34.43 mg LeucE/mL. The resulting liquor is rich in mineral salts (P, K, Ca, Mg, Na, Fe and Mn); its alcohol content is 45%. From a microbiological point of view, it is free of any pathogenic germ and meets the normative requirements. From the sensory analysis, it is well appreciated by all the tasters (>73%).

Keywords: *Ananas Comosus*; secondary metabolites; liquor, analyses; *Mondia whitei*

1. Introduction

Ananas Comosus is the second tropical fruit in world trade and contributes more than 20% to the world production of tropical fruits with 17 million tons [1]. In West Africa, Benin is the second largest producer of *Ananas Comosus* with 160,000 tons [1]. In 2015, Benin's share in world *Ananas Comosus* production was estimated at 315,795 tons, or 1.27% of world production. It contributes about 1.2% of GDP, and 4.3% of agricultural GDP [2]. Releases from the fruit processing industry are obtained every year in significant quantities all over the world [3]. *Ananas Comosus* canning industry generates 25 to 35% of waste per day [4]. One ton of fresh *Ananas Comosus* produces about 400 kg of waste, the pressing of which gives 260 to 300 liters of press juice [5]. Unfortunately in Benin, these rejects are not recycled and are often piled up in uncontrolled dumps located near processing centers [6]. Recent studies have been oriented towards the recovery of these fruit processing rejects. It appears that fruit wastes containing



fermentable sugars can no longer be abandoned in our environment, but they should be converted into useful food products such as wine, vinegar, ethanol, liquor etc.[7]. In Benin, the liquor is not only used during major festivals, ritual, religious and family ceremonies, but also on a daily basis as a sign of hospitality and respect. A good mastery of the transformation of *Ananas Comosus* by-products into a food drink would constitute an important issue, not only in the protection of the environment, but also at the socio-economic level. Despite this socioeconomic importance, little research has been conducted on the processing of *Ananas Comosus* suckers. It is in this logic that the present work aims at the valorization of the co-products of *Ananas Comosus* through the elaboration of the liquor reinforced by the active principles of *Mondia whitei*.

2. Materials and Methods

2.1 Materials It is made up of *Mondia whitei* root harvested in pobè (southern Benin) and co-products of *Ananas Comosus* collected in Akassato (southern Benin). The yeast strain used is “*Saccharomyces cerevisiae*” purchased on the local market, made in China, under the Angel brand.

2.2 Methods

2.2.1 Preliminary phytochemical screening

Secondary metabolites were carried out by coloration and precipitation reactions specific to each family of metabolites [8], [9], [10]

Table 1: Methods for identification secondary metabolites of *Mondia whitei* root

Secondary metabolites	Chemical test
Alkaloids	Mayer’s test and Dragendroff’s test
Anthocyanes	test with hydrochloric acid and ammonia
Anthraquinones	Borntranger’s test
Coumarins	Fluorescence at 365 nm
Flavanoids	shinoda test and magnesium powder
Tannins	stiasny test, Ferric chloride and sodium acetate test
Saponins	Frothing test
Leuco anthocyanins	Bate-Smith and metcalf
Mucilages	flaky test
Cyanogenic derivatives	picric acid test
Reducing compound	Fehling’s test
Sterols and terpenes	Liebermann-Burchard’s test
proteins	Biuret test

2.2.2 Technological diagram for making *Ananas Comosus* liquor

The liquor was obtained according to the method written by CEBENOR [11] and Hédiblé *et al.* [12]. Figure 1 below shows the production technology diagram of *Ananas Comosus* liquor.

2.2.3 Determination of phenolic compounds

- **Total phenol content:** Total phenolic content was determined using the Folin-Ciocalteu colorimetric method[13] with some modifications. This method consisted on using a mixture of phosphotungstic and phosphomolybdic acids, which were reduced during the oxidation of phenols into a mixture of tungsten blue oxide and molybdenum. Finally, the absorbance was measured at 760 nm using a spectrophotometer and the total phenol content are expressed in micrograms of gallic acid equivalence per mL ($\mu\text{gGAE/mL}$)[13], [14], [15].
- **Total flavonoids content:** The method of aluminum trichloride (AlCl_3) was used to quantify total flavonoids. This technique was based on the formation of aluminum complex flavonoids. The absorbance was read at 415 nm using a spectrophotometer and total flavonoid content are expressed in micrograms quercetin equivalence per mL ($\mu\text{gQE/mL}$)[16].
- **Condensed tannin content:** Vanillin and hydrochloric acid method was used to determine total condensed tannins content. The absorbance was measured at 500 nm using spectrophotometer and tannin content was expressed in micrograms catechin equivalence per mL ($\mu\text{gEC/mL}$) [17].



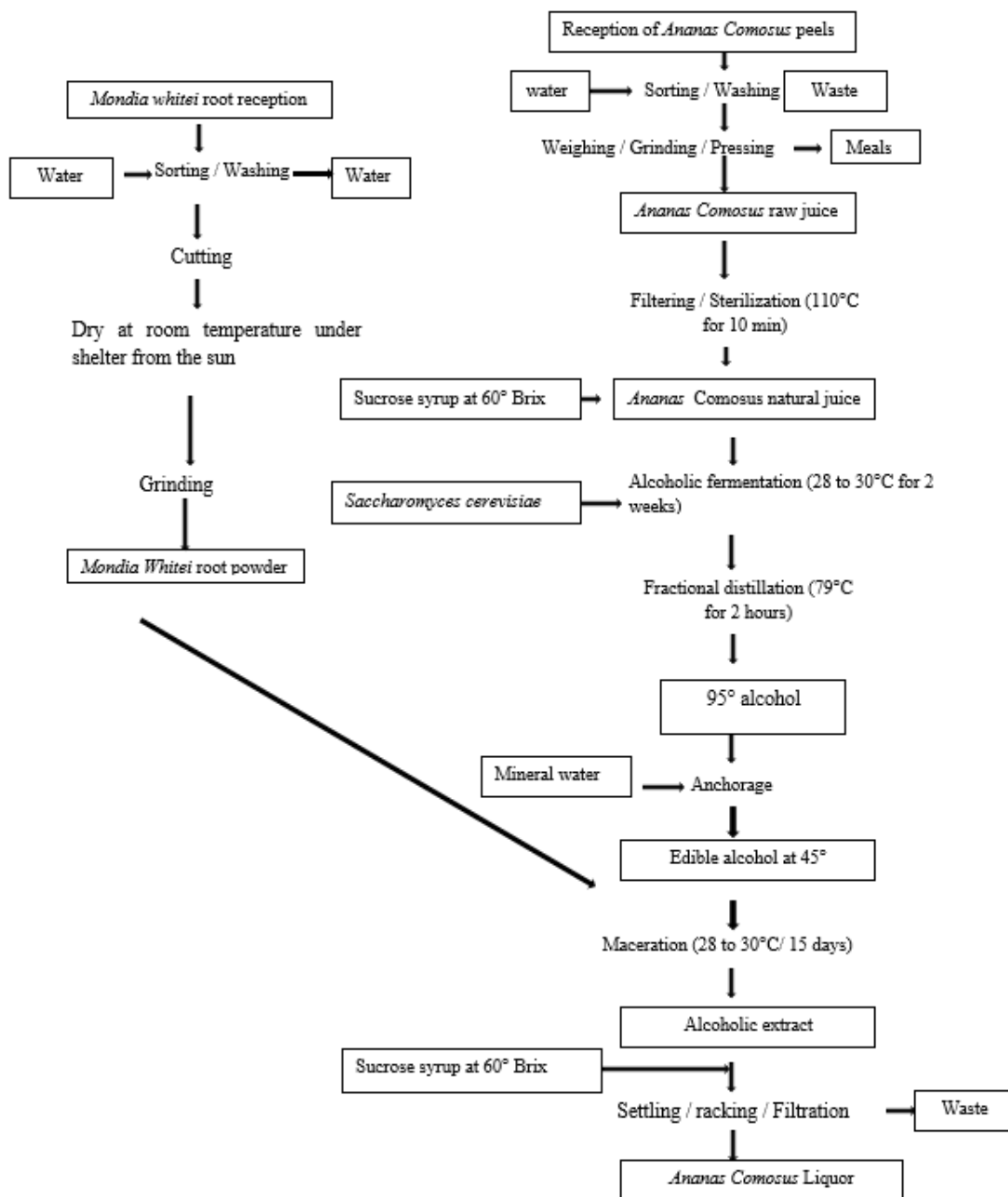


Figure 1: Technological diagram for making Ananas Comosus liquor

2.2.4 Minerals and Trace Elements Content: 5 grams of the sample were cremated following the procedure indicated above. The ash obtained was dissolved in 5 ml of hydrochloric acid (6N) and then evaporated on a hot plate at 125°C. The resulting residue was dissolved and recovered with HNO₃ (0.1M) in a 100 mL flask. This solution was used to obtain the content of the minerals by spectrophotometer Atomic Absorption. The results are expressed in relation to the dry matter [18].

2.2.5. Physicochemical analysis of formulated liquor: The pH was determined using a portable digital pH meter. The titratable acidity, expressed as percentage of citric acid, was determined by titration with NaOH (0.1N) in the presence of phenolphthalein. The soluble dry extract (°Brix) was determined using a portable refractometer of type (HI 96801 of 0-80° Brix. The relative density at 20°C was determined using a pycnometer according to the method

described by Massengo et al [19]. The dry matter was determined by drying in an oven at 105°C according to the recommended method [20]. The minerals were assayed by Atomic Absorption Spectrophotometry [18].

2.2.6. Microbiological analyzes: The microbiological quality of *Ananas Comosus* liquor was assessed through the determination of mesophilic aerobic flora, coliforms and yeasts and molds. 1 mL of the sample to be analyzed was taken aseptically to which 9 mL of sterile saline peptone water was added. The mixture was vortexed for 2 minutes, serving as the stock solution. Successive tenfold dilutions were prepared for seeding Petri dishes. The total aerobic mesophilic flora was counted using PCA agar (Plate Count Agar) at 30°C for 72 h. 1 mL of each of the dilutions as well as the stock suspension was introduced into different sterile Petri dishes. Then, 20 mL of the agar was poured into the contents of the Petri dishes. The whole was gently homogenized so as to incorporate the inoculum into the agar. After solidification, the agar is poured again to make a second layer. The count of germs was done according to the ISO 6222 1999 standard. The count of coliforms was carried out using VRBA (Violet Red Bile Agar) medium. 1 mL of each dilution was inoculated into a double layer of 10 to 15 mL of VRBA agar then incubated after solidification of the medium at 37°C for 72 h according to the ISO 21528-2: 2008 standard. Yeasts and molds were counted following incubation of 1 mL of each dilution on Malt Extract Agar (MEA, oxoid, CM 325, Hampshire, England) at 25°C for 72 hours. The count of germs in CFU/mL of samples analyzed was done according to the ISO 21527-1: 2008 standard.

2.2.6. Sensory analysis: Sensory tests were carried out at the end of production. From a panel of tasters, it was determined and evaluated the evolution of the sensory characteristics of the liquor produced according to the hedonic method [21]. The panel of tasters is made up of eighteen (18) people of different sexes and from all social categories, chosen from among the usual consumers of this product. The descriptors or attributes to consider are: color, flavor, aroma, smell, texture, bitterness, viscosity, taste and general impression.

3. Results & Discussion

3.1. Secondary metabolites identified from the root of *Mondia whitei*

The secondary metabolites of *Mondia whitei* are listed in Table 2. The root of *Mondia whitei* contains tannins, alkaloids, anthraquinones, saponosides, coumarins, flavonoids and reducing compounds. In the bark sample of *Mondia whitei* collected in Cameroon, reducing compounds, sterols and terpenes have been identified but note the absence of flavonoids [22]. At the level of *Mondia whitei* harvested in Zagnanando (Benin), tannins, flavonoids, alkaloids, coumarins and the absence of mucilage were identified [23]. In the sample of *Mondia whitei* collected in Congo, we note the presence of flavonoids, reducing compounds, saponins and tannins [24]. The variation in secondary metabolites observed in our samples compared to previous work could be related to the harvest period, the nature of the soil or climatic factors [25][26]. The presence of metabolites in the root of *Mondia Whitei* could give the formulated liquor antioxidant, vasculoprotective, antiallergic, anti-inflammatory, antidiarrheal, antimalarial properties [27], [28], [29], [30], [31], [32], [33], [34].

Table 2: Methods for the identification of secondary metabolites of root of *Mondia whitei*

Secondary metabolites	Present/Absent
Alcaloids	+
Tannins	-
Flavonoids	+
Anthocyanes	-
Leuco anthocyanins	+
Reducing compound	+
Mucilages	-
Coumarins	+
Cyanogenic derivatives	-
Saponosids	+
Anthraquinones	+
Sterols and terpenes	-

Legends: + : Present ; - : Absent

3.2. Physicochemical characteristics of *Ananas Comosus* liquor

Table 3 presents the values of the physicochemical parameters of *Ananas Comosus* liquor. According to this table, the pH, specific gravity and dry matter of pineapple liquor are respectively 4.75; 0, 97 and 12.22%. The titratable



acidity content of the liquor is 6.40 mg Eac/mL, with an alcohol content of 45% and a soluble solids content of 15° Brix. The pH of the formulated liquor is higher than that obtained on the red grape liquor in Portugal, which varies from 3.68 to 3.81[35]. The titratable acidity content of the liquor produced is well above that reported with the liquor obtained from cashew apple, the contents of which vary respectively from 0.6 to 1.0 mg Eac/mL [12]. The dry matter and soluble solids contents of the liquor produced are lower than those obtained by[12], whose values vary respectively from 6.65 to 24.69% and from 9.2 to 31.6° Brix. However, these values are lower than those obtained by Ferreira et al [35] ranging from 34.44 to 34.85% for dry matter and from 36.5 to 36.7 °Brix for soluble solids. The differences observed in these physicochemical parameters could be due to the degree of maturity of the raw material used.

Table 3: Physicochemical characteristics of *Ananas Comosus* liquor

Parametres	Ananas Comosus liquor
pH	4.75
Density	0.97
Dry matter content (%)	12.22
Titratable acidity (mgEac/ mL)	6.40
° Brix (%)	15.00
Alcoholic degree (%)	45.00

3.3. Mineral salt content

Table 3 below shows the mineral content of *Annas Comosus* liquor. From the analysis in Table 3, the *Annas Comosus* liquor produced has a high concentration of mineral salts. The contents of P, K, Na and Ca are respectively 60.00; 128.70; 7.96 and 17.44 mg/L. Its Mg concentrations; Fe and Mn of the liquor are of the order of 14.20; 1.57 and 0.14 mg/L. The contents of mineral salts and trace elements in the liquor from the Ciqual table (2020) [36] have a high content of sodium (72.50 mg/kg), magnesium (30.00 mg/kg) and potassium (300.00 mg/ kg) compared to those found in our work. On the other hand, the iron (0.80 mg/kg) and calcium (10.00 mg/kg) contents are lower than those obtained in the *Annas Comosus* of liquor produced. These differences could be due to the raw material used.

Table 4: Mineral salt content

MSC	Ca	Fe	Na	Mg	K	P	Mn
(mg/Kg)	17.44	1.57	7.96	14.20	128.70	60.00	0.14

MSC: mineral salt content ; K: Potassium ; Ca: Calcium ; Na: sodium; P: phosphorus; Mg: Magnesium ; Fe: Iron ; N : nitrogen

3.4. Content of phenolic compounds in the formulated liquor

Table 5 presents the content of phenolic compounds in the liquor produced. This table shows that the liquor contains a significant amount of tannins, flavonoids and total phenols. The total phenol concentration is 9.49 mgEAG/mL. The contents of total flavonoids and condensed tannins are respectively 6.05 µg EQ/mL and 119.5 mgELeuc/mL. The total phenol content obtained from our liquor is lower than the values reported by Ferreira et al[35] between 24.70 and 25.70 mgEAG/mL. On the other hand, it is higher than those obtained by Hédible[12] ranging from 0.26 to 0.55 mgEAG/mL. Indeed, flavonoids from food plants have been reported to provide biological benefits, such as reducing the risk of cancer and cardiovascular disease [37].

Table 5: Content of phenolic compounds in the liquor produced

Phenolic compound	Total phenol content (µgGAE/mL)	Total flavonoids content (µgQE/mL)	Condensed tannin content (µgCE/mL)
	9.49	6.05	119.5

3.5. Microbiological quality

Table 6 presents the results of the microbiological analysis of the liquor produced



From the microbiological point of view (table5), the liquor produced does not contain coliforms. As for yeasts and moulds, they are below the microbiological criteria. These values are in accordance with the beverage standard which indicates that the number of yeasts and molds must not exceed 10^3 CFU/mL [38]. The count of the total aerobic mesophilic flora is less than 10^3 CFU/mL. These values meet the normative requirements. Thus, the *Annas Comosus* liquor produced is of acceptable microbiological quality.

Table 6: Microbiological characteristics of *Annas Comosus* liquor

Microorganisms	Liquor	Normative values
TAMF	10^2 UFC/ mL	$< 10^3$ UFC/mL (FSD, 2018)
Total coliforms	Absent	$< 10^2$ UFC/ mL (FSD, 2018)
Yeasts	4.10^1 UFC/ mL	$< 10^3$ UFC/ mL [38]
Molds	10^1 UFC/mL	$< 10^3$ UFC/ mL [38]

FSD: Food Security Division; TAMF: Total Aerobic Mesophilic Flora; CFU: Colony Format Unit

3.6. Sensory analysis

Table 7 presents the averages obtained at the end of the sensory tests carried out on the *Annas Comosus* liquor. As for the general impression, this figure shows that the tasters really liked the *Annas Comosus* liquor with an average appreciation rate of 73.38%. Parameters such as color, bitterness and viscosity of *Annas Comosus* liquor with a respective appreciation rate of 68.80%, 62.77% and 67.22% are moderately liked by tasters. In addition, the alcoholic flavor and the sweet aroma of the elaborate liquor having received an average appreciation of 70.55% and 70% are much appreciated by tasters.

Table 7: Sensory analysis of *Annas Comosus* liquor

Parameters	Appreciation rate
Color	68.8
Alcoholic flavor	70.55
Sweet aroma	70
Bitterness	62.77
Viscosity	67.22
General impression	73.38

Legends: 10: I extremely hated; 20: I hated it a lot; 30: I slightly hate; 40: I'm indifferent; 50: I liked it a little; 60: I moderately liked; 70: I loved; 80: I really liked

4. Conclusion

This study made it possible to produce and characterize the liquor from the co-products of *Ananas Comosus* on the physicochemical, microbiological and sensory levels. The phytochemical screening showed that the root of *Mondia whitèi* contains secondary metabolites such as tannins, flavonoids, alkaloids, reducing compounds, anthraquinones, saponins and coumarins. The product produced has an acid pH and high levels of dry matter, titratable acidity and soluble dry matter. The evaluation of phenolic compounds shows that they are rich in total phenols, flavonoids and condensed tannins. Mineral analysis indicates that the liquor contains a significant amount of mineral salts; essential to man. From a microbiological point of view, it is free of any pathogenic germ and meets the normative requirements. From the sensory analysis, it appears that it is well appreciated by all the tasters (>73%). The transformation of *Ananas Comosus* co-products into liquor solves problems related to pollution and is of undeniable economic importance.

References

- [1]. Agbangba, C., E., Dagbenonbakin, D., G., Houssou, P., Assea, D., E., Sossa, E., L., Kotomalè, U. A., Ahotonou, P., Ndiaga, C., & Akpo, L.E., (2015). Influence de la fertilisation minérale sur la qualité physico-chimique et organoleptique du jus d'ananas transformé de Cayenne lisse au Bénin. *Int. J. Biol. Chem. Sci.* 9(3), 1277-1288;
- [2]. Kpenavoun, S., C., Gandonou, E., et Fiogbe N. (2017). Technical efficiency of small-scale pineapple production in Benin. Published by EDP Sciences.
- [3]. Grigoras, 2012. Valorisation des fruits et des sous-produits de l'industrie de transformation des fruits par extraction. ICOA - Institut de Chimie Organique et Analytique.



- [4]. Py C., et Tisseau M., A., (1965). L'ananas. Collection technique agricoles et production tropicales. G-P Maisonneuve et Larose; Paris. 289p;
- [5]. Estanove P., (1980). Fabrication du vinaigre à partir du jus de presse de déchet de conserverie d'ananas, Institut de recherches sur les fruits et les agrumes.
- [6]. Aboh, A.,B., Ehounsou, M.,A., Olaafa, M. & Brun, A. (2008). Complémentation alimentaire des ovins Djallonké avec les sous-produits de transformation d'ananas: potentiel nutritif, préférence et développement pondéral. Bulletin de la Recherche Agronomique du Bénin, 61 : 25-30.,
- [7]. Kaidi F., Touzi A., 2001. Production de bioéthanol à partir des déchets de dattes. Review of renewable energy, 75-78;
- [8]. Dohou N., Yamni K., Tahrouch S., Idrissi Hassani L.M., Badoc A., Gmira N. (2003). Screening phytochimique d'une endémique ibéro-marocaine, *Thymelaea lythroides*. Bulletin de la société pharmaceutique de Bordeaux, 142: 61-78;
- [9]. Koudoro Y.,A., Awadji J.,M., Botezatu D.,A., Olaye T. , Agbangnan D. C. P., Alitonou G., A., Avlessi F., Dinica R., M., and Sohounhloue C., K. D., (2021). Phytochemical analysis, antioxidant and anti-inflammatory activities of *Chassalia kolly* leaves extract, a plant used in Benin to treat skin illness. GSC Biological and Pharmaceutical Sciences, 15(03), 063–072;
- [10]. Agbangnan D.C.P., Tachon C., Bonin H., Chrostowska A., Fouquet E. and Sohounhloue C.K.D., (2012). Phytochemical study of a tinctorial plant of benin traditional pharmacopoeia: The red sorghum (*sorghum caudatum*) of Benin. Scientific Study & Research, vol. 13(2), pp: 121-135.
- [11]. CEBENOR, 2005. Norme béninoise NB 03.04.001. Alcool de manioc à usage alimentaire: Spécifications. Bénin, 5 p.;
- [12]. Hédiblè L., G., (2018). Valorisation agroalimentaire des pommes d'anacarde (*Anacardium occidentale* L.) acclimatées au centre du Bénin : Etude des conditions optimales de bioproduction d'alcool alimentaire et des liqueurs. Thèse de soutenance pour l'obtention du grade de Docteur de l'Université d'Abomey-Calavi, 172 p;
- [13]. Lupoae P., V. Cristea, D. Borda, M. Lupoae, G. Gurau, Dinica RM (2015). Phytochemical Screening: Antioxidant and Antibacterial Properties of *Potamogeton* Species in Order to Obtain Valuable Feed Additives. Journal of Oleo Science, 1-13;
- [14]. V.L.Singleton, R. Orthofer, R.M. Lamuela-Raventos, Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. Methods Enzymol, 1999, 299: 152-178;
- [15]. Mbacke D.I.S., A.D.Fall , K. Diatta-Badji, A. Sarr, S. Madieye, S. Moussa, A. Mbaye, W. Diatta, Bassene Evaluation de l'activité antioxydante des extraits hydro-ethanoliques des feuilles et écorces de *Piliostigma thonningii* Schumach. Int. J. Biol. Chem. Sci, 2017, 11(2): 768-776;
- [16]. Djeridane Y., Yous M., B. Nadjemi, D. Boutassouna, P. Stocker, N. Vidal (2006). Antioxidant activity of some Algerian medicinal plants extracts containing phenolic compounds. Food Chem, , 97: 654-660;
- [17]. Heimler D.,Vignolini P., Giulia M., Francesco V.F., (2006). Rmani A Antiradical activity and polyphenol composition of local Brassicaceae edible varieties. Food Chemistry, 99: 464-469;
- [18]. Kanninkpo C., (2013). Synthèse de quelques méthodes d'analyse par le Laboratoire des Sciences du sol, des Eaux et Environnement (LSSEE/CRA Agonkanmey/INRAB) pour les produits végétaux ; Rédigé sur la base des documents de travail; 2p.,
- [19]. Massengo V., Loumouamou B.W., Diakabana P., Silou T. (2014). Ethanol production fermentation of the pulpe of BOKO mango. International Journal of Chemical Science and Technology,4:71- 77;
- [20]. AOAC, 2000. Official Methods of Analysis. 17 th Edn., Association of Official Analytical Chemistry, Arlington, Virginia, Gaithersburg, MD, USA.
- [21]. Watts B.M., Ylimaki G.L., Jeffery L.E. et Elias L. G., 1991. Méthode de base de l'évaluation sensorielle des aliments. 159p;
- [22]. Watcho Pierre, Patrick Brice Deeh Defo, Modeste Wankeu-Nya, Miguel Carro-Juarez, Telesphore Benoît Nguelefack and Albert Kamanyi (2013). *Mondia whitei* (Periplocaceae) prevents and *Guibourtia tessmannii* (Caesalpiniaceae) facilitates fictive ejaculation in spinal male rats. BMC Complementary and Alternative Medicine. 13(4) ;pp:1-9;
- [23]. Gbohaida Virginie., (2013). Etude cinétique de l'extraction des polyphénols naturels et bioactifs de quatre plantes médicinales du Bénin : *Jatropha multifida*, *Carapa procera*, *Mondia whitei* et *Pterocarpus erinaceus*. Mémoire DEA p 52



- [24]. Mbadu Z., Ntumba M., Sumba F., Benandwenga M., Ekalakala T., 2015. Contrôle de la qualité microbiologique et physicochimique de la boisson artisanale Londo à base de *Mondia whitei* (Hook. f.) Skeels) (Apocynaceae). p 39-40;
- [25]. Daddona P.E., Wright J.L., Hutchinson C.R. (1976). "Alkaloid catabolism and mobilization in *Catharanthus roseus*", *Phytochem*, 15, pp. 941-945.
- [26]. Manolaraki F. (2011). "Propriétés anthelmintiques du sainfoin (*Onobrychis viciifoliae*). Analyse des facteurs de variations et du rôle des composés phénoliques impliqués. Thèse de Doctorat de l'université de Toulouse III, Toulouse;
- [27]. Bruneton, J. *Pharmacognosie, phytochimie, plantes médicinales*, 2ème édition, Paris : Editions médicales internationales, Tec et Doc Lavoisier, (1993). P 915;
- [28]. Christina, A.J.M., M.A. Jose, S.J.H. Robert, R. Kothai, N. Chidambaranathan and P.Muthuman, (2003). Effet of *Indigofera aspalathoides* against Dalton's ascitic lymphoma. *Fitoterapia*, 74(3):280-283;
- [29]. Elliott M. J., Chithan K., Theorides T. C. (2000). The effects of plant flavonoids on Mammalian cells: implications for inflammation, Heart disease, and Cancer. *Pharmacol. Rev.* 52(4), pp:673-7;
- [30]. Fenglin H., Ruilin L., Bao H., Liang M.; (2004). Free Radical scavenging activity of extracts prepared from fresh leaves of selected Chinese medicinal plants. *Fitoterapia* 75(2), pp:14-23;
- [31]. Nacoulma O. G. (1996). *Plantes médicinales et Pratiques médicales Traditionnelles au Burkina Faso: cas du plateau central T1&T2*. Thèse Doct. d'Etat ès Sciences Nat. Université de Ouagadougou, 242 et 285;
- [32]. Narayana K., Reddy M. S., Chaluvadi M. R., Krishina D.R., (2001). Bioflavonoids classification, pharmacological, biochemical effects and therapeutic potential. *Indian. J. of pharmaco.*, 33, 2-16;
- [33]. Ueda H., Yamazaki C., and Yamazaki M. (2002). Luteolin as an anti-inflammatory and anti-allergic constituent of *Perilla frutescens*. *Biol.Pharm. Bull.*, 25(9) , pp: 1197- 1202;
- [34]. Yenjai C., Prasanphen K., Daodee S., Wongpanich V., Kittakoop P. (2004). Bioactive Flavonoids from *Kaempferia Parviflora*. *Fitoterapia*, 75, 89-92.
- [35]. Ferreira A., M., Sousa I., Ricardo-da-Silva J. M. (2001). Liqueur des raisin, optimisation de sa formulation. Caractérisation physico- chimique et sensorielle. Universidade Técnica de Lisboa, Instituto Superior de Agronomia, Laboratório Ferreira Lapa (Sector de Enologia) Lisboa, Portugal. *Ciência Tèc. Vitiv.* 16 (2), pp:65-79;
- [36]. Ciqal, 2020. Nutrition database for standard reference, release 7, 01. Department of Nutrition, National Food Institute, Technical University of Denmark version 7, 1, 2009;
- [37]. Brito E.S., Araujo M.C.P., Lin L.Z., Hamly J. (2007). Determination of the flavonoid components of cashew apple (*Anacardium occidentale*) by LCDAD- ESI/MS. *Food Chemistry*, 105: 1112-1118;
- [38]. Luxembourg, 2011. Critères microbiologiques applicables aux denrées alimentaires et les lignes directrices pour l'interprétation. Ministère de la santé, Grand-Duché de Luxembourg, 49p.

