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Research Article

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Role of Ginger as Bioenhancer in the Treatment of Diabetes

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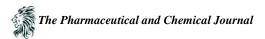
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Abstract: Diabetes is a common and very prevalent disease and a major health problem worldwide. It has been reported to be the major cause of blindness, kidney failure, lower-extremity amputation cardiovascular diseases and premature mortality. Diabetes has increasing cases in rural and poor populations throughout the world. The possibility of its management by the oral administration of hypoglycemic agents has stimulated great research interest in over the years. Considering that diabetes mellitus is a heterogeneous disorder with multiple causes, the combination of therapeutic agents aimed at specific patho-biological pathways of diabetes and its complications may result in a better and more effective management of this disorder. The present study was conducted to evaluate the anti-diabetic activity of Glibenclamide in combination with Ginger. Diabetes was induced by administration of Streptozotocin (i.p 65 mg/Kg). Biochemical parameters were evaluated in the study. When Glibenclamide is used in combination with Ginger, even at sub therapeutic level of Glibenclamide showed similar effects as that of therapeutic dose of Glibenclamide.

Keywords: Diabetes, Glibenclamide, Streptozotocin, Biochemical

Introduction

Diabetes mellitus (DM) is a metabolic disorder of multiple etiologies characterized by chronic hyperglycemia (high blood sugar) with disturbances in carbohydrate, fat and protein metabolism resulting from defects in insulin secretion, insulin action or both [1]. Diabetes is a common and very prevalent disease affecting the citizens of both developed and developing countries. The greatest increase in prevalence is however expected to occur in Asia and Africa, where more patients will likely be found by 2030 [2]. Diabetes has increasing cases in rural and poor populations throughout the world, despite major investigation into understanding the pathophysiology and treatment of diabetes mellitus. It has continued to be a major health problem worldwide. The possibility of its management by the oral administration of hypoglycemic agents has stimulated great research interest in over the years. Though different types of oral hypoglycemic agents are available along with insulin for the management of diabetes mellitus, there is increased demand by patients for the use of herbal preparations with hypoglycemic activity [3]. The increasing prevalence of this disorder not only poses severe medical implications but also has financial consequences due to the costs of managing this disorder and its associated complications. Considering that diabetes mellitus is a heterogeneous disorder with multiple causes, the combination of therapeutic agents aimed at specific pathobiological pathways of diabetes and its complications may result in a better and more effective management of this disorder [4]. Even though the currently available drugs may be valuable in the management of diabetes mellitus, these drugs have limitations due to undesirable adverse effects such as hypoglycemia, weight gain, secondary failure, and inability to arrest pancreas degeneration or diabetic complications which have been linked to oxidative



stress [5]. Several classes of allopathic drugs namely Sulphonylureas, Biguanides, Thiazolidinediones, α -glucosidase inhibitors and semi synthetic Insulin are extensively used to control the hyperglycemia. These drugs exert different modes of action to reduce the elevated blood glucose, but none of them were found to be effective in providing complete glycemic control in diabetic patients, moreover these drugs also exert their individual adverse effects such as pregnancy related complications and hypoglycaemic episodes when administered to the diabetic patients [6]. Hence there is a need for an antidiabetic drug/formulation that can provide optimal glycemic control without producing unwanted side effects in the host system [7].

Materials and Methods

Experimental Animals

Healthy Wistar albino rats (150-200 g) of either sex were used for the experiments. They were maintained under standard conditions (temperature 22 ± 2 °C, relative humidity $60\pm5\%$ and 12 h light/dark cycle). The animals were housed in sanitized polypropylene cages containing sterile paddy husk as bedding. They had free access to standard pellet diet and water ad libitum. Experiments were conducted between 10:00 to15:00 h. All experimental protocols will be reviewed and approved by the Institutional Animal Ethical Committee (IAEC) prior to the initiation of the experiment and the care of the laboratory animals will be taken as per the CPCSEA regulations.

Chemicals

All the chemicals and reagents used were of analytical grade and were purchased from YARROW CHEM, LOBA CHEM, HIMEDIA and Agappe diagnostics.

Preparation of Aqueous Extract of Ginger

Aqueous Ginger extract was prepared from locally available Ginger rhizomes. Ginger rhizomes (500g) were peeled on crushed ice and were cut in to small pieces and homogenized in 750 ml cold, sterile 0.9% NaCl solution and 250 ml ice cold water to make the volume 1000 ml. The homogenization was carried out in a blender for 12 minutes. The homogenized mixture was filtered three times through cheese cloth. The filtrate was centrifuged at 2000 rpm for 10 min and the clear supernatant fraction was separated and volume made up to 1000 ml with normal saline. The concentration of this Ginger preparation was considered to have 500mg/ml on the basis of the weight of the starting material according to the formula. The extract was stored in sample tubes at -4 °C until fed to rats [8].

Anti-Diabetic Activity

Streptozotocin induced anti-diabetic activity [9,10]

Fasting blood glucose was determined after depriving food for 16 h with free access to drinking water. Hyperglycemia was induced by single i.p injection of 65 mg/kg of STZ in citrate buffer, freshly prepared and injected within 5 minute of preparation to prevent degradation. After administration of STZ the animals had free access to feed and water ad libitum. The development of hyperglycemia in rats was confirmed by fasting blood glucose estimation 48 h post STZ injection wherein animal were fasted overnight again for blood collection from tail vein. The rats with fasting blood glucose level of above 200 mg/dl at 48 h after STZ injection were considered diabetic and included in the study.

Citrate buffer (pH 4.4, 0.1 M)

Citric acid monohydrate 0.6306 g was dissolved in 50 ml of distilled water. Trisodium citrate 0.7352 g was dissolved in 25 ml of distilled water. 28 ml of Citric acid monohydrate and 22 ml solution were taken and mixed together. It is made up to a volume of 1000 ml with distilled water. The pH of the solution was adjusted to pH 4.4.

Experimental Design

Animals will be randomly divided into 4 groups of 6 each and assigned as below. Group I: Vehicle control (Citrate buffer).



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Group II: Diabetic control (Streptozotocin 65mg/Kg). Group III: Diabetic + Glibenclamide (1mg/Kg) Group IV: Diabetic + Glibenclamide (0.5 mg/Kg) + Ginger (500mg/Kg)

The parameters studied were as follows:

- Biochemical parameters such as
 - i. Fasting blood glucose
 - ii. Serum glutamic pyruvate transaminase (SGPT)
 - iii. Serum glutamic oxaloacetate transaminase (SGOT)
- Endogenous antioxidant parameters includes
 - i. Superoxide dismutase (SOD)
 - ii. Catalase (CAT)

Estimation of Biochemical Parameters

The above treatment was carried out in each group of animals for 21days.

Fasting blood glucose was measured using select simple single touch glucometer. Blood samples were withdrawn under mild anesthesia from tail tip of the overnight fasted animals on 1st, 7th, 14th, and 21st day. On 21st day the blood was collected by retro orbital puncture and the serum was obtained by centrifuging the blood samples at 3000 rpm for 10 min and they were used for estimation of SGPT, SGOT by using a corresponding kit from Agappe Diagnostics Pvt. Ltd and the intensity of the coloured complex formed after treating with these reagents were estimated in semi-auto analyzer.

Estimation of Antioxidant parameters

Tissue preparation

Animals were sacrificed by cervical dislocation and were perfused transcardially with an ice-cold saline. The whole liver was perfused in situ with ice-cold saline, dissected out, blotted dry and immediately weighed. A liver homogenate was prepared with ice-cold saline-EDTA. The homogenate was centrifuged at 10,000 rpm for 10 min and the pellet discarded The supernatant was again centrifuged at 20,000 rpm for 1 hour at 40°C.

Superoxide Dismutase [11]

2.8 ml of sodium carbonate buffer (0.05 mM) and 0.1 ml of tissue homogenate or sucrose (Blank) was incubated at 300C for 45 minutes. Then, the absorbance was adjusted to 0 to sample. Thereafter, the reaction was initiated by adding 10 μ l of adrenaline solution (9mM). The change in absorbance was recorded at 480nm for 8-12 minutes. Throughout the assay, the temperature was maintained at 300C. The results are expressed as unit (U) of SOD activity per mg of protein.

Catalase [12]

The liver homogenates containing $5\mu g$ total protein was mixed separately with $700\mu l$, 5mM hydrogen peroxide and incubated at $37^{\circ}C$. The disappearance of peroxide was observed at 240nm for 15min. One unit of catalase activity is that which reduces $1\mu mol$ of hydrogen peroxide per minute.

Statistical analysis

Results of biochemical estimation were reported as mean \pm S.E.M. The total variation present in a data was analyzed by one way analysis of variance (ANOVA). P value less than 0.05 was considered as statistically significant.

Results and Discussion

Effect on Fasting Blood Glucose Level Fasting blood glucose (FBG) level was within the range of 80-90 mg/dl in all the groups prior to STZ administration. Treatment with STZ in normal saline (65 mg/kg, i.p) had increased the FBG



Table 1: Serum Glucose in STZ Induced Diabetic Rats							
Groups	Blood glucose level (mg/dl)						
	Before diabetic induction	Day 0	Day 7	Day 14	Day 21		
Normal control	85.62±1.38	86.10±1.86	86.34±1.92	87.37±2.56	85.25±3.16		
Diabetic control	85.08±4.58	298.45 ± 2.26	315.72 ± 5.36	311.87±4.56	321.91±5.23		
Glibenclamide	84.93±1.132	295.16 ± 2.56	244.21±6.54**	189.59±4.89**	127.36±3.45**		
Glibenclamide +	85.406±4.67	304.08 ± 7.56	268.56±4.32**	192.45±5.92**	140.46±4.67**		
Ginger							

level above 290 mg/dl after 48 h. Treatment with Glibenclamide and Glibenclamide in combination with Ginger significantly normalized the elevated blood glucose level as shown in Table 1.

Values are mean \pm SEM (n=6 except in control) one way ANOVA followed by Dunette's test. Where ** represents highly significant at p < 0.01

Estimation of Serum Biomarkers

After 21 days of experiment, serum biomarkers such as SGPT and SGOT level were significantly elevated in diabetic control group. In animals treated with Glibenclamide and Glibenclamide in combination with Ginger, SGPT and SGOT levels were decreased significantly (p < 0.001, p < 0.01 respectively) as compared to the diabetic control (Table 2).

Table 2: SGPT and SGOT Levels in Diabetic Rats					
Groups	SGPT	SGOT			
Normal	58.61±2.65	62.63±3.75			
Diabetic	139.48 ± 2.38	$148.72{\pm}1.05$			
Glibenclamide	79.52±3.54***	76.57±1.95**			
Glibenclamide +Ginger	92.80±2.11***	84.64±1.69**			

Antioxidant Parameters

From antioxidant studies, it was found that STZ induced diabetic control animals showed a significant decrease in the levels of SOD and CAT as compared to normal control. Standard group treated with Glibenclamide and test group treated with Glibenclamide + ginger combination showed significant increase in CAT and SOD as compared to diabetic control (Table 3).

Table 5: Serum Antioxidant Parameters in Diabetic Rats					
Groups	SOD	Catalase			
Normal	11.36 ± 3.25	6.45±2.57			
Diabetic	3.28 ± 1.78	2.35±1.15			
Glibenclamide	9.52±2.54***	4.46±2.75**			
Glibenclamide + Ginger	8.46±1.32***	4.05±2.96**			

Discussion

The present study was undertaken to evaluate the anti-diabetic activity of a standard synthetic anti-diabetic drug Glibenclamide in combination with an herbal anti-diabetic drug Ginger in diabetic rats.

In the study, the blood glucose levels in STZ treated rats were significantly increased as compared to normal rats where as the group treated with standard drug Glibenclamide (1mg/Kg) and those treated with Glibenclamide (0.5mg/Kg)+ginger combination showed significant reduction in glucose levels. The results indicate that the combination Glibenclamide and Ginger could lead to increase in the effect of Glibenclamide that may be helpful to reduce the dose of Glibenclamide and to minimize the adverse effects as well as maintain enhanced therapeutic hypoglycemic effect.



Insulin deficiency leads to various metabolic aberrations in the rats; the rise in blood glucose level is accompanied by increase in SGPT and SGOT level. In this present study, Glibenclamide and Glibenclamide + ginger combination treated serum reports showed significant reduction in SGPT and SGOT level which indicate that combination therapy with sub therapeutic dose of Glibenclamide and Ginger is comparable to effect of Glibenclamide at its therapeutic dose.

Oxidative stress plays a major role in the pathogenesis of both types of diabetes mellitus. Free radicals are formed in diabetes by glucose oxidation, protein glycation, and the subsequent degradation of glycated proteins [13]. High levels of free radicals and the simultaneously declined antioxidant enzyme levels lead to cell damage, inactivation of enzymes and lipid peroxidation [14]. Superoxide dismutase and catalase play an important role in the detoxification of super oxide anion and H_2O_2 respectively. In present study, Catalase and SOD which are most important antioxidant enzymes were found to be decreased in diabetic control group with induction of diabetes, but treatment with Glibenclamide and Glibenclamide + ginger combination level of both enzymes were found to be increased. Glibenclamide + Ginger combination treatment showed more increased levels of SOD and Catalase than that of Glibenclamide treatment [15].

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